Please amend the application as follows:

In the specification:

Please replace the paragraph at page 1, lines 9 through 15, with the following:

This application is a continuation of U.S. Application No. 09/025,178, filed February 18, 1998 (now abandoned), which claims the benefit of U.S. Provisional Application No. 60/038,059, filed February 18, 1997 and U.S. Provisional Application No. 60/066,288, filed November 25, 1997, the contents of which are incorporated herein by reference in their entirety.

Please replace the paragraph at page 19, lines 2 through 17, with the following:

Spleens were removed from mice 10 days after the last injection. The spleens from 3-10 mice in each treatment group were pooled. Single-cell suspensions were prepared by grinding tissue through a sterile nylon mesh. Erythrocytes were removed by suspending the cells in pH 7.2 lysis buffer (0.15 M NH_4Cl, 1 M $KHCO_3$, 0.1 mM Na_2EDTA) and rinsing the cells two times with RPMI 1640 medium. Splenocytes were then cultured at 1 x 10^7 cells/ml in 96-well round bottom microculture plates in RPMI 1640, supplemented with 10% FCS and 50 μM 2-ME at 37°C in 5% CO_2. The cells were stimulated with recombinant ovalbumin (10 μg/ml), SIINFEKL peptide (SEQ ID NO: 1) (10 μg/ml) or with Con A (5 μg/ml). Cell culture supernatants were removed at 72 h. A sandwich ELISA using paired monoclonal antibodies (Endogen, Cambridge, MA) was used to measure IFN-γ.

Please replace the paragraph at page 19, lines 19 through 26, with the following:

Single-cell suspensions of splenocytes were prepared as above. 25 x 10^6 splenocytes were cultured with 5 x 10^6 irradiated (15,000 rads) E.G7-OVA cells in RPMI 1640 supplemented with 10% FCS, 50 μM 2-ME, 1 mM sodium pyruvate and 100 μM non-essential amino acids. After 6-7 days in culture, splenocytes were purified by FICOLL-PAQUE (Pharmacia, Piscataway, NJ) density centrifugation and then utilized as effector cells.

Please replace the paragraph at page 22, lines 13 through 34, through page 23, lines 1-12, with the following:

Whether mice injected with soluble protein without adjuvant could be primed to produce anti-ovalbumin T cells was investigated (Figure 1A). C57BL/6 mice were inoculated i.p. with 120 pmoles of ovalbumin (ova) or with 120 pmoles of ovalbumin-hsp70 fusion protein (ova-hsp70) in PBS. A second equivalent dose was given s.c. at two weeks. A third group of mice was injected with 120 pmoles of ovalbumin-p24 gag fusion protein (ova-p24), purified as described in Suzue and Young, *J. Immunol. 156*:873-879 (1996)), in order to examine the immune responses elicited by administering ovalbumin covalently linked to a protein other than hsp70, in the absence of adjuvant. Splenocytes of immunized mice were removed ten days after the s.c. immunization and for each mouse group, 5-10 spleens were pooled and splenocytes from immunized mice were cultured *in vitro* for 6 days with irradiated E.G7-OVA cells (syngeneic EL4 cells transfected with ovalbumin) without added interleukins (Moore, M.W. *et al.*, *Cell*, *54*:777-785 (1988)). The cultured cells were then used as effector cells in CTL assays. Cells from mice injected with ovalbumin protein or with ovalbumin-p24 fusion protein were unable to lyse T2-K^b target cells or T2-K^b cells pulsed with SIINFEKL peptide (SEQ ID NO: 1). In contrast, effector cells from mice primed with ovalbumin-hsp70 fusion protein were able to lyse T2-K^b cells pulsed with SIINFEKL peptide (SEQ ID NO: 1). See Figure 1A, where the splenocyte cultures derived from mice immunized with ova ▫, ova-p24 ▽ and ova-hsp70 ■, which were used as effector cells in a standard cytotoxicity assay, are shown. ^{51}Cr-labeled target cells were used: T2-K^b cells (--) and T2-K^b pulsed with SIINFEKL peptide (SEQ ID NO: 1)(—) at 300 μg/ml.